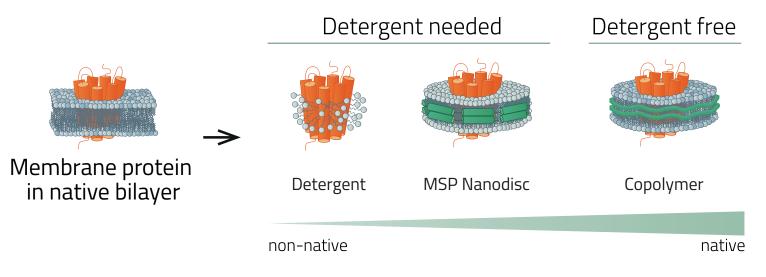
NativeMPTM Platform Next Generation Copolymers for the Characterization of Membrane Proteins in Near-Native Conditions

Philipp Hanisch, Lea Esser, Felipe Merino, Jan Kubicek and Barbara Maertens

Abstract: The study of membrane proteins has been revolutionized by the development of next-generation copolymers, offering profound advancements in purification. Membrane proteins, and represent approximately 60% of current drug targets. Research on these proteins is essential to combat a broad range of diseases, including cardiovascular disorders, neurological conditions, and cancers. Traditional detergents, while useful, often strip away essential lipids, leading to protein instability and loss of function. In contrast, copolymers utilized in Cube Biotech's unique NativeMPTM platform maintain a lipid-rich environment that present a comprehensive workflow, from gene to structure, utilizing next-generation polymers for full-length membrane protein purification. We demonstrate the efficacy of this approach with a case study on an ion channel, showcasing the retention of functional lipids and the maintenance of protein folding and expression in a suitable host, followed by solubilization with our Native MPTM platform. These copolymers enable the extraction of membrane proteins in a lipid environment that closely mimics their native conditions. Subsequent steps include purification, functional integrity and are amenable to high-resolution structural determination. This approach represents a significant leap forward in membrane protein research, offering a viable alternative to detergent-based methods. By preserving the lipid milieu, next-generation copolymers facilitate the study of membrane proteins in a state that closely resembles their physiological context. This enables more accurate insights into their mechanisms and interactions, paving the way for advancements in drug discovery. Our findings underscore the potential of copolymer-based techniques to transform the landscape of membrane protein characterization and functional analysis, thereby enhancing our ability to develop targeted therapies for a wide range of diseases.

1. Introduction Part One

Stabilizing Membrane Proteins

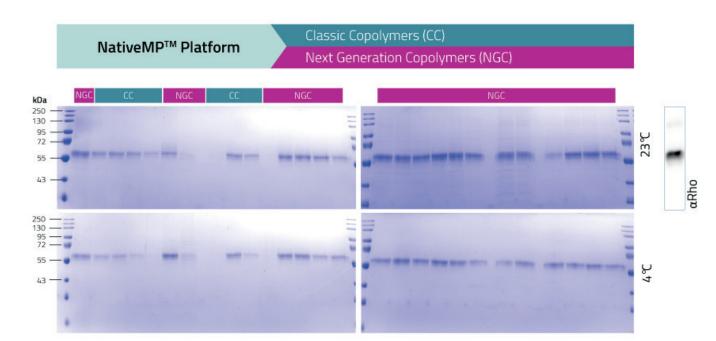


- Loss of lipids
- lipid-nanodisc complex. -> Not optimal
- lipids still attached. -> Best option we have in vitro

2. Screening is Key

The NativeMP™ Platform vs Detergent

Working at 23°C significantly boosts elution quantity (up to 200%) compared to 4°C



temperatures

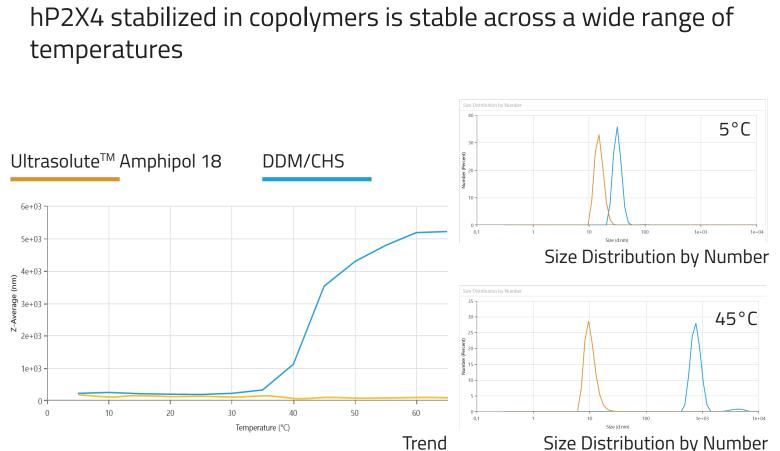


Fig. 2A: Full-length hP2X4-Rho has been solubilized and stabilized from cell membrane in one step at 4°C/ 23°C with classical as well as next generation copolymers (bold). Protein of interest was purified at 4°C/23°C from solubilizate via Rho-MagBeads, eluates were analysed via SDS-Page and Western-Blot (A). Working at 23°C significantly boosts elution quantity (up to 200%) compared to 4°C.



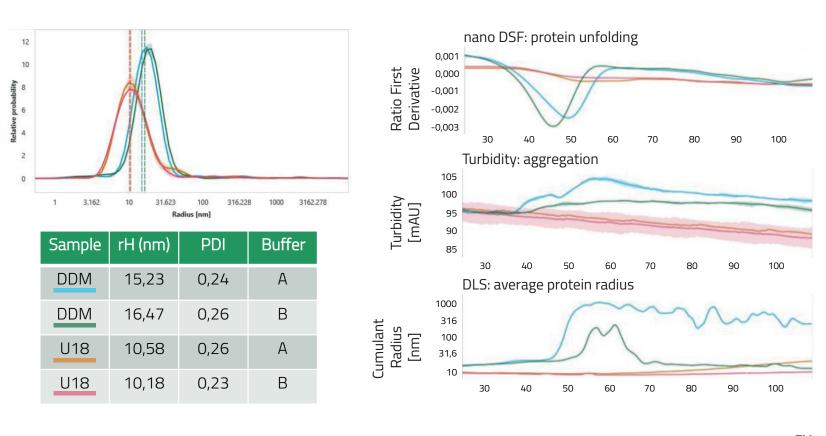


Fig. 2B: DLS measurements & nDSF analysis with Prometheus Panta of NativeMP[™] stabilized hP2X4 (Ultrasolute[™] Amphipol 18 picked as representative Copolymer) & hP2X4 stabilized DDM/CHS. Elution in two different buffers directly from Rho-MagBeads (B). The DDM/CHS micelle increased the hydrodynamic radius (rH) of the measured particles compared to the Native MP[™] disc. All eluates show a comparable polydispersity index (PDI). hP2X4 stabilized in DDM/CHS has been purified as described in Kotov et al., 2019.

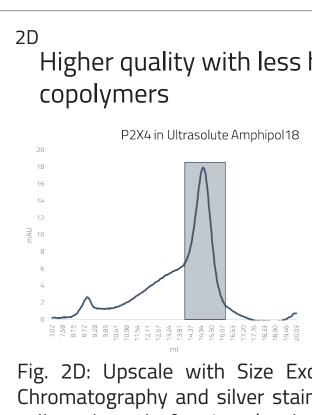
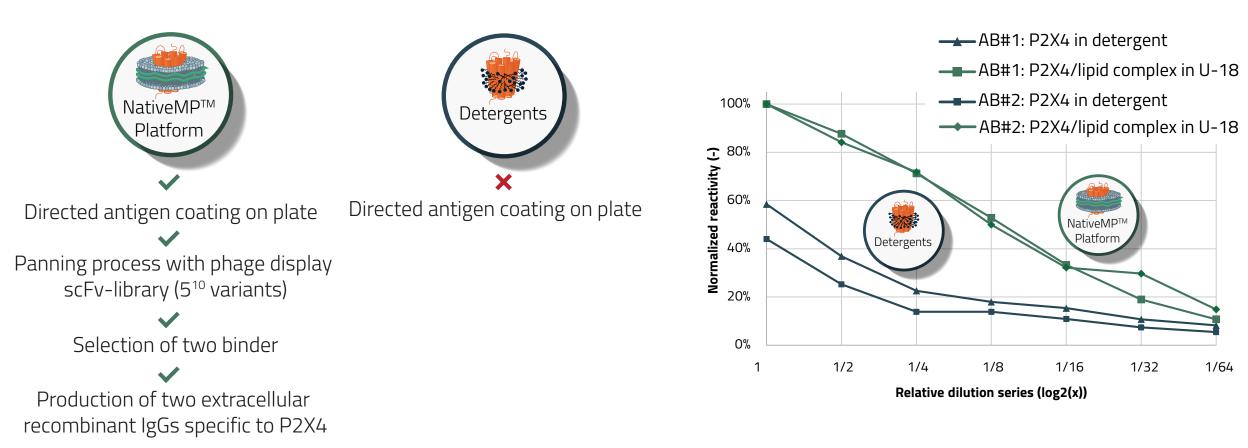


Fig. 2D: Upscale with Size Exclusion Chromatography and silver staining of P2X4 1µg ^{Ultrasolute™DDM/} collected peak fraction (peak 1 for DDM/CHS) of NativeMP[™] stabilized hP2X4 (Ultrasolute[™] Amphipol 18) and hP2X4 stabilized in DDM/CHS. The detergent stabilized hP2X4 is less pure --compared to the Native MP[™] stabilized ____ sample even after careful peak selection. NativeMP[™] stabilized hP2X4 purified at 23°C results in a ~400% hP2X4 increase compared to DDM/CHS stabilized hP2X4 at 4°C.

3. Making the Impossible Possible **Copolymer Stabilized P2X4 Enables Antibody Generation**



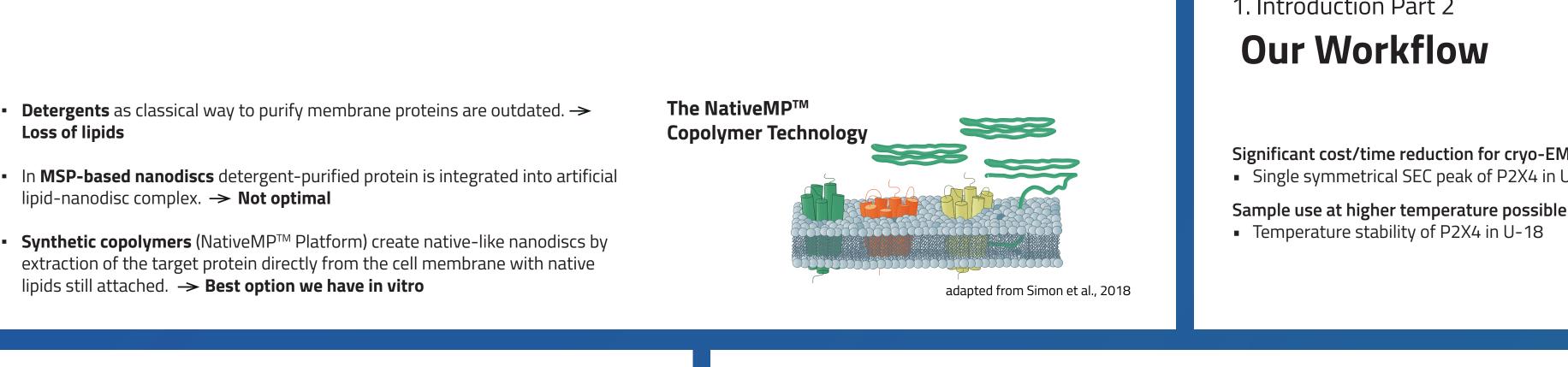


Fig. 2C: NativeMP[™] stabilized hP2X4 (Ultrasolute[™] Amphipol 18) is highly stable, shows no unfolding or aggregation over a temperature ramp from 4°C to >60°C (only DLS data shown). hP2X4 stabilized in DDM/CHS shows an inflection point at 42-45°C in nDSF and DLS measurements (C).

Higher quality with less hands-on time with

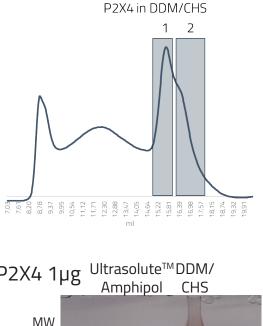
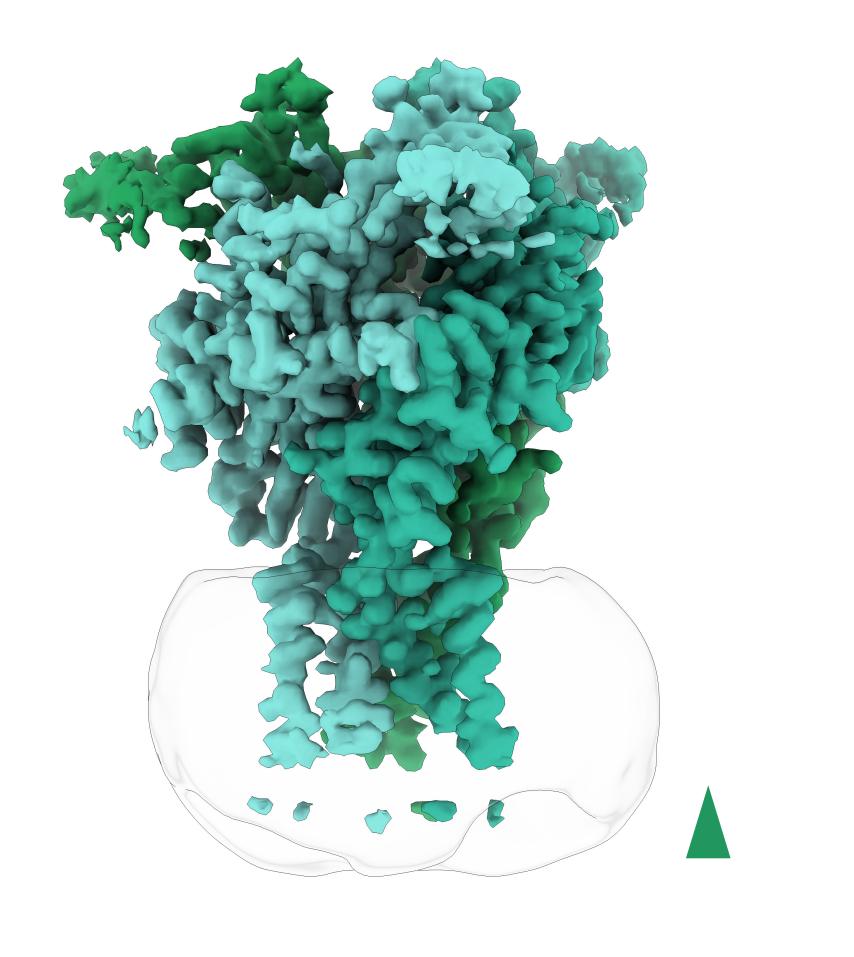




Fig. 3: Antibody generation in collaboration with Proteogenix: Generation of highly specific antibodies against hP2X4 utilizing a scFv-library in phage Antibody display. Generation was only possible in NativeMP[™] **stabilized hP2X4.** Specific IgGs showed >2x increased binding to Ultrasolute[™] Amphipol 18 (U-18) stabilized hP2X4 compared to DDM/CHS stabilized hP2X4.

Finding The Perfect Match hP2x4 Full Length Structure



Cube Biotech

6. Finding The Perfect Match **Structure Determination**

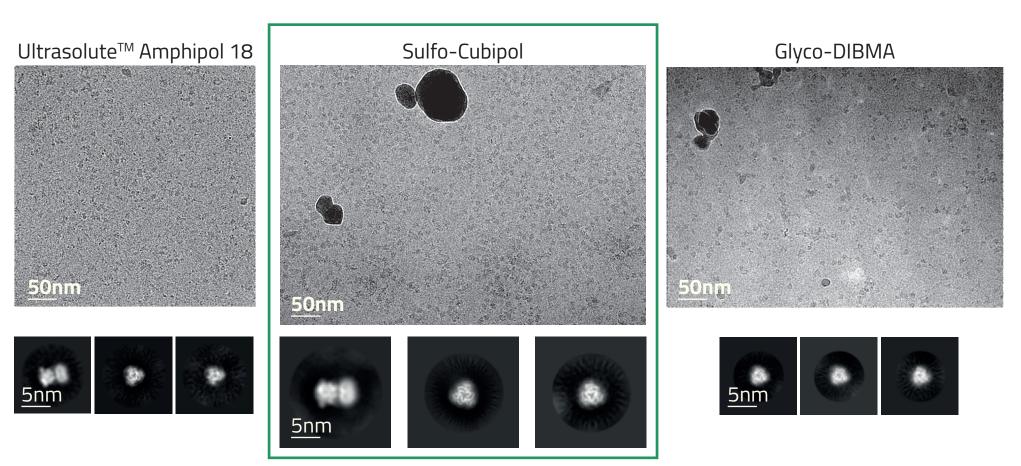
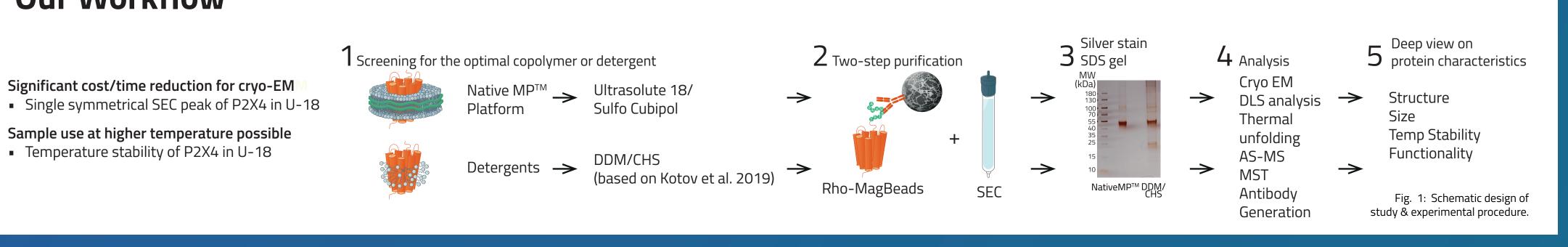


Fig. 6: Careful copolymer selection and an extensive screening process is the key to success when trying to obtain optimal results in cryo-EM. Sulfo-Cubipol produces an optimal particle orientation distribution for hP2X4. Early refinement cycles already produce a reconstruction of < 3.5 Å global resolution.

1. Introduction Part 2 **Our Workflow**



4. Yet Again Impossible in Detergent – Possible in NativeMP™ First Ever Ion-Channel Ligand Screened in AS-MS

Validation of method using **reference ligands** to P2X4 in copolymer Preserving lipid environment ensures **native-like ligand binding**

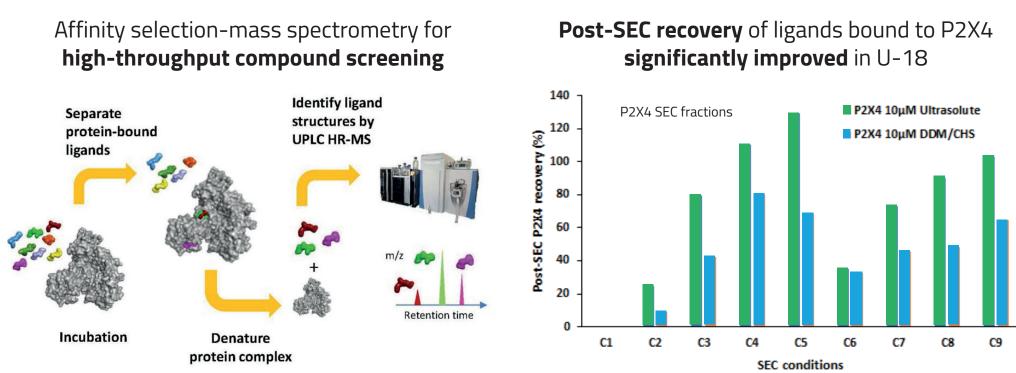


Fig. 4: Copolymer stabilized P2X4 enables Antagonist Screening with AS-MS. AS-MS antagonist screening in collaboration with Edelris: hP2X4 stabilized in U-18 or DDM/CHS was tested in 9 different SEC conditions for AS-MS approach. Post-SEC recovery of ligands bound to hP2X4 significantly improved in U-18. AS-MS ligand screening was only possible in NativeMP[™] stabilized hP2X4. kD of three reference ligands could successfully be measured with U-18 stabilized hP2X4.

5. Reproduction in Method Variety Copolymer enables Antagonist Screening with MST/ Spectral Shift

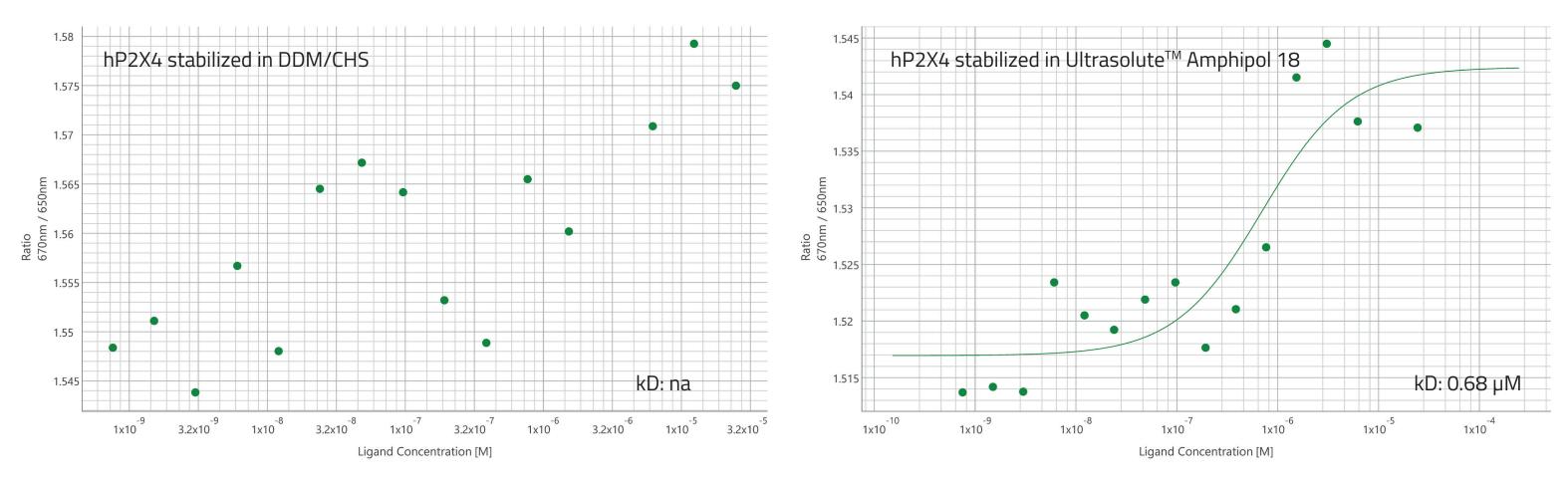


Fig. 5: hP2X4 stabilized in U-18 or DDM/CHS was labeled with a 647-NHS dye and mixed with increasing amounts of 5-BDBD. The binding affinity was measured in solution using a Nanotemper Monolith and standard capillaries after binding check and buffer optimization. A kD measurement was only possible when hP2X4 was stabilized in synthetic copolymer (U-18, kD: 0.68 µM). DDM/CHS stabilized hP2X4 showed no measureable kD when mixed with 5-BDBD.

7. Conclusion

Universal Applicability Beyond hP2X4

Copolymers have a broad range of advantages compared to classic detergents. The NativeMP[™] Platform is the way to go if you want to purify active, stable and correctly folded membrane proteins in their native lipid environment. We show that the NativeMP[™] Platform beats detergent in hands-on time, efficiency, temperature stability, functionality, storability, ligand interaction and antibody generation. Copolymers are the future of membrane protein science and the drug targeting market.

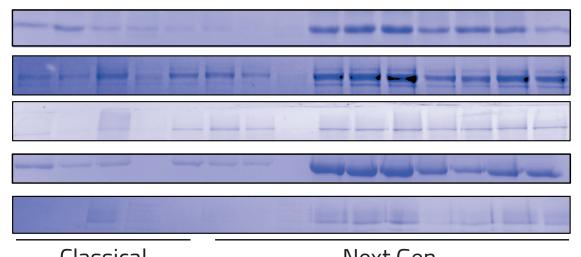


Fig 7: 5 important Drug Target MPs screened, purified and 2D classified in less than 4 weeks.

Classical Copolymers

Next Gen Copolymers



Reference ligands tested **successfully**

Ligand	Literature IC ₅₀	Measured K _d
BX 430	0.54 µM	0.60 µM
5-BDBD	0.75 µM	0.85 µM
Indophagolin	2.71 µM	3.35 µM

References 1. Kotov, V., Bartels, K., Veith, K. et al. High-throughput stability screening for detergent-solubilized membrane proteins. *Sci Rep 9*, 10379 (2019). https://doi.org/10.1038/s41598-019-46686-8